Aspidophoroides monopterygius (Bloch, 1786) Agonidae Alligatorfish

Myomeres:

Vertebrae:

Dorsal fin rays:

Pectoral fin rays:

Pelvic fin rays:

Caudal fin rays:

Anal fin rays:

Meristic Characters

48 - 52

48-52

5-6

5-6

10 - 11

I. 2

5 + 5

Range: Arctic to northern Pacific and Atlantic oceans; in the western North Atlantic Ocean from Greenland to Cape Cod, widespread on Grand Bank, northern and western Gulf of Maine, northeast Georges Bank; rarely occurring as far south as New Jersey

- **Habitat**: Benthic in depths of 18–332 m over sandy or muddy substrates, also pebbly bottoms or those with broken shell; larvae have been collected in brackish as well as highly oceanic waters
- Spawning: Not well described; larvae present in Gulf of Maine and Salem Harbor (Massachusetts) during spring

Eggs:	- Undescribed	; those of oth	ner species dei	mersal, large, de	eposited in clump	os under rocks
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Larvae: – Body very elongate, with preanus length <50% SL

- Head elongate, with pointy snout, somewhat concave profile, mouth only reaching anterior edge of eye
- Series of spines on top of head; see checklist below
- Note constriction, or 'loop', in mid-gut, with anus and terminus of gut semi-trailing
- Pectoral fin early forming, large and fan-shaped
- Dorsal and anal fins short-based, opposite each other; single dorsal fin situated at the halfway point of body
- Caudal fin small and rounded
- Rows of tiny spines form along the body; these correspond to rows of plates (modified scales) in adults
- Pigmentation includes a characteristic series of blotches along the body, ending on the caudal fin base; pectoral fin rays tipped with black pigment; melanophores scattered over much of gut; patches of fine pigment on edge of preopercle

Head Spine Checklist:

Parietal:	two rows of small spines in preflexion larvae become arched crest in later larvae
Coronal:	spine present
Tympanic:	spine present
Superior infraorbital:	1 st SIO present
Preopercular:	usually 4 large spines along margin; 2nd and 3rd spines form first; 1st added in late flexion
Interopercular:	spine present (not shown in key)
Supracleithral:	spines present after late flexion
Subopercular:	spines present after late flexion
Nasal:	spine present in postflexion
Dentary:	spine present in postflexion (not shown in key)
Supraocular:	spine present in postflexion
Pterotic:	spine present in postflexion
Postocular:	spines situated on top of head in postflexion; note shape as dermal plates

- Note:
- 1. Pectoral fin ray count lower than in Uncina olriki
 - 2. Larvae occur in plankton until they settle to bottom habitats at a size of about 29.0 mm

Figures: Adult: Kanayama, 1991; A, D: Bev Vinter (Busby, 1998); B: Bigelow and Schroeder, 1953; C: Wayne Laroche (Washington *et al.*, 1984b); agonid head spines: Busby and Ambrose, 1993

References: Bigelow and Schroeder, 1953; Elliott and Jiminez, 1981; Washington *et al.*, 1984a; Yabe, 1985; Maeda and Amaoka, 1988; Busby, 1998; Klein-M^{ac}Phee, 2002h

Aspidophoroides monopterygius



Leptagonus decagonus (Bloch and Schneider, 1801) Agonidae Atlantic poacher



Damaa	Anotic coop into conthe	m Desife and Atlantic account in the most with	[]		
Kange:	Atlantic from arctic w Scotia and Grand Bank,	vaters to Greenland, Newfoundland, northern Nova , as far south as 44°15'N	Meristic Characters Myomeres: 44–49		
Habitat:	Benthic in deep, cold w muddy substrates	waters (0-5°C) in depths of 27-392 m over sandy or	Vertebrae: 44–49 Dorsal fin rays: V–VIII, 5–8 Anal fin rays: 6–8		
Spawning:	Not well described; rip have been collected spri	e eggs have been found winter into late spring; larvae ing into late summer	Pectoral fin rays: 15–17 Pelvic fin rays: I, 2 Caudal fin rays: 6+5		
Eggs:	 Undescribed; those of under rocks 	of other species demersal, large, deposited in clumps			
Larvae:	 Body very elongate, with preanus length <50% SL Preanus length decreases through development; distance between anus and anal fin origin increases Body compressed in tail region; not as hexagonal in cross-section as that of <i>Ulcina olriki</i> Head elongate, with pointy snout, somewhat concave profile, mouth only reaching anterior edge of eye Series of spines on head; see checklist below Barbels form under maxillary during larval stage Note constriction, or 'loop', in mid-gut, with anus and terminus of gut semi-trailing Pectoral fin early forming, large and fan-shaped; pelvic fin rays form late Dorsal (2) and anal fins short-based; 2nd dorsal fin origin posterior to level of anal fin origin Caudal fin rounded Rows of tiny spines form along the body; these correspond to rows of plates (modified scales) in adults Pigmentation includes characteristic series of blotches along body, ending at tip of notochord; pectoral fins covered with scattered melanophores, may have darker margin; most gut pigment on dorsal surface; some pigment 				
Head Spine	Head Spine Checklist: (See key to head spines on Asnidanharaides manantervaius figure page)				
iicuu Spiiic	Parietal:	row of small spines in early flexion, form ridges or cres ion larvae	ts with serrate edges in late flex-		
	Preopercular: Supraocular: Nasal: Superior infraorbitals: Inferior infraorbitals: Frontal: Postocular: Posttemporal: Pterotic:	usually 4 large spines along margin; all spines present b spines develop by 16.3 mm spines develop by 16.3 mm spines present in juveniles spines present in juveniles spines present in juveniles spines present in juveniles; note shape as dermal plates spines present in juveniles spines present in juveniles	y 13.7 mm		
Note:	1. This species has 2 c area have only one	lorsal fins, the anterior of which consists of spines; the of dorsal fin and lack dorsal fin spines	ther 2 species that occur in study		

- 2. High number of pectoral fin rays
- 3. Larvae occur in plankton until about 24 mmSL; size at which they settle to the bottom unknown

Figures:Adult: D. R. Harriott (Scott and Scott, 1988); A, B, D: Bev Vinter (Busby, 1998); C: Ehrenbaum, 1905References:Dannevig, 1919; Dunbar, 1947; Washington *et al.*, 1984a; 1984b; Yabe, 1985; Maeda and Amaoka, 1988; Busby, 1998

Leptagonus decagonus



B. 15.7 mmSL



C. 21.0 mmSL



D. 29.5 mmSL

Ulcina olriki (Lütken, 1876) Agonidae Arctic alligatorfish

Frontal:

Pterotic:

Postocular:

Posttemporal:



Range:	Northern Pacific and Atlantic ocea tic to coastal waters of Labrador a	ns; in the western North Atlantic from Arc- s far south as 52°22'N; also Hudson Bay	Meristic Characters		
Habitat:	Benthic in depths of 18–110 m ov	er sandy, muddy or rocky substrates	Myomeres: 37–39 Vertebrae: 37–30		
Spawning:	Not well described; 250 ripe eggs Greenland	found in a single female during Jul off west	Dorsal fin rays: 5–7 Anal fin rays: 5–7		
Eggs:	 Ripe ovarian eggs 0.75–1.5 mm sal, large, deposited in clumps u 	in diameter; those of other species demer- nder rocks	Pectoral fin rays: 14–15 Pelvic fin rays: I, 2 Caudal fin rays: 5+5		
Larvae:	- Body very elongate, with pream	us length <50% SL			
	- Body is laterally compressed an	d hexagonal in cross-section			
	- Head elongate, with pointy snow	it, somewhat concave profile, mouth only reach	ing anterior edge of eye		
	- Series of spines on head; see ch	ecklist below			
	 Note constriction, or 'loop', in m 	hid-gut, with anus and terminus of gut semi-trail	ing		
	- Pectoral fin early forming, large and fan-shaped				
	- Dorsal and anal fins short-based, opposite each other; single dorsal fin situated at halfway point of body				
	– Caudal fin small and rounded	 Caudal fin small and rounded 			
	 Rows of tiny spines form along the body; these correspond to rows of plates (modified scales) in adults; spines more prominent than in larvae of <i>Leptagonus decagonus</i> 				
	 Pigmentation includes a characteristic series of blotches along the body, ending on the caudal fin base; pectoral fin rays tipped with black pigment; melanophores scattered over much of gut, especially mid-section and termi- nus; pigment extends from bar on body onto dorsal and anal fins; scattered spots on head and snout 				
Head Spine Checklist: (See key to head spines on Aspidophoroides monopterygius figure page)					
	Parietal:	row of small spines in early flexion, form ridge	es or crests in late flexion		
	Preopercular:	usually 4 large spines along margin; all spines	present by 13.7 mm		
	Supraocular:	spines developed by 16.3 mm			
	Nasal:	spines developed by 16.3 mm			
	Superior and inferior infraorbital:	spines present in juveniles			

1. Pectoral fin ray count higher than in Aspidophoroides monopterygius Note:

2. Larvae occur in plankton until about 20-21 mmSL; size at which they settle to the bottom unknown

spines present in juveniles; note shape as dermal plates

spines present in juveniles

spines present in juveniles

spines present in juveniles

Figures: Adult: Kanayama, 1991; A, D: Bev Vinter (Busby, 1998); B: Rass, 1949 (redrawn); C: Dunbar, 1947 **References**: Dunbar, 1947; Washington et al., 1984b; 1984b; Yabe, 1985; Maeda and Amaoka, 1988; Busby, 1998

Ulcina olriki



D. 25.0 mmSL

Cyclopterus lumpus Linnaeus, 1758 **Cyclopteridae** Lumpfish

- Arctic and North Atlantic oceans; in the western North Atlantic from Range: Davis Strait and Greenland to Chesapeake Bay
- Pelagic, becoming demersal during reproduction in depths to 329 m Habitat:
- Spawning: Early spring in coastal waters
- Eggs: - Demersal, adhesive, pink, in masses
 - Deposited in nests constructed by male
 - Diameter: 2.2-2.6 mm
 - Chorion thick, hard
 - Oil globules: multiple, coalesce into one
 - Incubation 10-70 days; male provides protection from predation and aeration
- Larvae: - Hatching occurs at 5.6 mmSL or smaller (Newfoundland) with pectoral rays and suction disk fully formed
 - Hatching size 4.0-7.4 mm in Gulf of Maine
 - Tadpole-shaped with large, wide head, slender tail
 - Tubercles begin to form over eyes at 18 mm, over body at 22 mm; by 25 mm, small tubercles scattered over much of body
 - Full complements of 2nd dorsal and anal fin rays formed by 5.6 mmSL; 1st dorsal and caudal fin rays complete by 6.1 mmSL
 - 1st dorsal fin pedunculate
 - Pigmentation includes scattering of small spots over most of head and body; note unpigmented band from snout to opercle edge;

Note: 1. Larvae and juveniles may use tide pools as nursery from Jun-Dec, often associated with algae and seagrass

Early Juvenile:



H. 34.0 mm

Note development of tubercles, large, pedunculate 1st dorsal fin and slender form; background pigment is green, olive, brown or yellow; have occurred in near-surface waters in Bay of Fundy Jul-Sep

Meristic Characters			
Myomeres:	28–29		
Vertebrae:	28-29		
Dorsal fin rays:	VI–VIII, 9–11		
Anal fin rays:	9–10		
Pectoral fin rays:	19–20		
Pelvic fin rays:	Disk		
Caudal fin rays:	11-12		

Cyclopterus lumpus



Liparis tunicatus Reinhardt, 1837 Liparidae Kelp snailfish



Meristic Characters			
Myomeres:	45-50		
Vertebrae:	45-50		
Dorsal fin rays:	39–44		
Anal fin rays:	33-37		
Pectoral fin rays:	32-38		
Pelvic fin rays:	Disk		
Caudal fin rays:	4-6+4-6		

- **Range**: Arctic Ocean from western Alaska and Chukchi Sea to western Greenland; in study area occurs south along the coast of Labrador to Saglek Bay (58°30' N, 63°00' W)
- **Habitat**: Demersal in shallow water (usually <50 m), close to shore, often associated with kelp beds (*Laminaria* sp.); uses suction disk to attach to kelp blades; bottom substrates usually pebbly or rocky, more rarely sandy to muddy
- **Spawning**: Undescribed; a single report of undeveloped gonads during summer (Able and M^cAllister, 1980)
- Eggs: Undescribed
- Larvae: Developmental series undescribed; characters based on a single lateflexion stage larva (Fig. A)
 - Head flat-topped
 - Nostrils tubular, prominent
 - Suction disk moderately large (13-14% SL)
 - Gill slit opening very small throughout development; see comparative table on L. fabricii page
 - Head length 22-27% SL
 - Preanus length about 41% SL
 - Snout to anal fin 46–48% SL
 - Fin rays complete by 17.0 mm
 - Pigmentation includes scattering of melanophores over dorsal and lateral surfaces of head and body, with few blotches on dorsal fin rays; ventral surfaces of head and body relatively unpigmented
- Note: 1. Similarities to larvae of *L. coheni* and *L. gibbus*:
 - High numbers of myomeres and fin rays
 - Large sizes at all developmental stages
 - No development of notch in anterior part of dorsal fin
 - 2. Best distinguished from *L. gibbus* by lower pectoral fin ray count and very small gill opening; this species and *L. coheni* do not overlap geographically



A. 17.9 mmNL

Figures: D. R. Harriott (Scott and Scott, 1988); A: M. P. Fahay (Able *et al.*, 1986); B: Betsy Washington (Able *et al.*, 1984)
References: Able and McAllister, 1980; Able *et al.*, 1984; Scott and Scott, 1988

Liparis tunicatus

Notes on Larvae of Cyclopteridae and Liparidae

Larvae of these families typically have a short, bulbous head and tadpole-shaped body. They are similar to larvae of some cottids, scorpaenids, triglids and ceratioids, but they:

- Lack head spines
- Have a restricted gill opening that closes ontogenetically; (see table on Liparis fabricii page for rates of closing)
- Have a highly modified pelvic disk that forms early in ontogeny (may be secondarily lost in some genera)
- Have a broad-based pectoral fin that begins at the upper level of eye and ends near cleithral symphysis
- Have a well-developed finfold that envelopes the posterior end of the body



B. 5.0 mmSL (*Cyclopterus lumpus*)

Larvae of Liparidae are similar to those of Cyclopteridae, but they:

- Have more myomeres (38-86 vs: 23-29)
- Are more elongate
- Have more dorsal and anal fin elements
- Are less developed and less pigmented at hatching

Larval characters in 3 genera of Liparidae

	Liparis	Careproctus	Paraliparis
Suction disk	Present, forms early	Present, forms early	Absent
Hatches	In preflexion stage	In late flexion or postflexion	In late flexion or postflexion
Nostrils	2 pairs	Single pair through development	Single pair through development
Lower lip	Typically tri-lobed	Single	Single
Pectoral fin base	Cluster of spots on inner surface	Inner surface not pigmented with cluster of spots	Inner surface not pigmented with cluster of spots
Fin rays	Begin anteriorly	Complete complements at hatch	Complete complements at hatch
Head length	Longer	Shorter	Shorter
Preanus length	Longer	Shorter	Shorter

Larvae of *Rhodichthys* are undescribed; they hatch from large eggs (5.5 mm diameter) and lack modified pelvic suction disks (Andriashev, 1954). *Psednos* species are humpbacked dwarves, seldom exceeding 50 mmSL, and lack suction disks. Their larvae are also undescribed.

Liparis fabricii Krøyer, 1847 Liparidae Gelatinous snailfish



,	
Meristic Charac	ters
Myomeres:	48-53
Vertebrae:	48-53
Dorsal fin rays:	43–49
Anal fin rays:	36-42
Pectoral fin rays:	32-37
Pelvic fin rays:	Disk
Caudal fin rays:	4-5+5-6

- **Range**: Circumpolar; in study area from Arctic and Atlantic coastal Canada from northern Ellesmere Island to Grand Bank
- Habitat: Pelagic in midwater layers and benthic in depths >50 m to a maximum 600 m
- Spawning: Sep–Oct in Russian Arctic (Andriashev, 1954); larvae present Aug near Baffin Island and Labrador (Dunbar, 1947)

Eggs: – Undescribed

- Larvae: Relatively shallow-bodied, short-headed form with large eye, short preanus length, and small suction disk
 - Relatively large sizes at all developmental stages (hatching, flexion, disk formation)
 - Disk forms at 8.6-11.5 mm
 - Flexion begins at lengths of 11.9-13.4 mm and is complete by 48.2-52.1 mm
 - Relatively high number of myomeres and fin rays
 - Nostrils complete split into 2 pairs at 14.1–17.2 mm
 - Pectoral fin rays complete during postflexion; caudal fin rays complete by 21 mm; dorsal and anal fin rays complete by about 20 mm
 - Darkly pigmented peritoneum and pectoral fin; prominently barred pigment pattern forms on body and fins; acluster of small melanophores on the medial surface of the pectoral fin base; at preflexion, larvae have scattered row of melanophores along base of anal fin and on dorsolateral surface of gut
- Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids
 - 2. The larvae of this species, *L. atlanticus* and *L. gibbus* are the only liparid larvae with lateral body pigment present during preflexion stage
 - 3. The gill slit closes ontogenetically in *Liparis* larvae; the rate of closing is less in *L. fabricii*; *L. tunicatus* has a very small opening throughout development (see table below)

Species	Flexion Stage	Gill Slit Length (% HL)
Liparis atlanticus	Preflexion	64
	Postflexion	13
Liparis inquilinus	Preflexion	66
	Postflexion	20
Liparis coheni	Preflexion	60
	Postflexion	19
Liparis gibbus	Preflexion	45
	Postflexion	36
Liparis fabricii	Preflexion	51
	Flexion	43
Liparis tunicatus	Flexion	13
	Postflexion	9

Figures:Adult: Stein and Able, 1986; A: M. P. Fahay (Able *et al.*, 1986); B–D: Betsy Washington (Able *et al.*, 1984)References:Dannevig, 1919; Able and McAllister, 1980; Able *et al.*, 1986

Liparis fabricii



C. 16.7 mmNL (Ventral View) Venter of gut densely pigmented

Note very small suction disk





Liparis coheni Able, 1976 Liparidae Gulf snailfish



Myomeres:

Vertebrae:

Dorsal fin rays:

Pectoral fin rays:

Pelvic fin rays:

Caudal fin rays:

Anal fin rays:

Meristic Characters

42-46

42 - 46

36-41

30-35

32-40

Disk

5+5-6

Range:	Western North Atlantic Ocean from Gulf of St. Lawrence to Gulf
	of Maine and Georges Bank

- Habitat: Demersal in depths of 4–210 m
- Spawning: Winter to early spring in St. Lawrence River Estuary and Gulf of Maine
- Eggs: Undescribed
- Larvae: Hatching size undescribed; smaller than 7.0 mm
 - Early larvae deep-headed with tapering tail; later larvae become less tapered with shallower heads
 - Preanal ventral outline tends to be flat
 - Suction disk becomes visible at 5.4 mm (or earlier)
 - Eye diameter to disk length ratio smaller than in comparably sized L. gibbus
 - Nostril splits into 2 pairs at 7.8-9.6 mm
 - Preanus length increases from 44% SL in preflexion to 47% SL in flexion; decreases during postflexion to 43% SL
 - Snout to anal fin increases from 46% SL during preflexion to 48% SL during postflexion
 - Head length increases from about 24% SL during preflexion to 29% SL during postflexion; head length larger than in comparably sized *L. gibbus*
 - Flexion occurs between 8.5 and 20.7 mm
 - Sequence of fin ray formation: $P_1 D, A C$
 - Notch does not form in anterior part of dorsal fin; fin rays are uniform or graduated in length
 - Pigmentation includes a cluster of small melanophores on the medial surface of the pectoral fin base; at preflexion, larvae have melanophores on dorsolateral surface of gut and on dorsum of head; small melanophores form clusters on ventral surface of head and posterior to disk; melanophores along base of anal fin occur on every 2nd or 3rd anal fin pterygiophore; scattered spots on caudal fin rays; lateral surface of body without pigment until >12.0 mm
 - Adult pigmentation is attained during late flexion or postflexion stages; ranges from uniformly dark to a variety
 of striped or barred patterns
- Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids
 - 2. Similarities to larvae of L. gibbus and L. tunicatus:
 - High numbers of myomeres and fin rays
 - Large sizes at all developmental stages
 - No development of notch in anterior part of dorsal fin

Liparis coheni



E. 20.0 mmNL

Liparis gibbus Bean, 1881 Liparidae Dusky snailfish



Meristic Characters			
Myomeres:	44–50		
Vertebrae:	44–50		
Dorsal fin rays:	38–46		
Anal fin rays:	32-37		
Pectoral fin rays:	37–45		
Pelvic fin rays:	Disk		
Caudal fin rays:	4-5+5-6		

- Range:Circumpolar; in western North Atlantic from Canadian Arctic and
Greenland to southern Gulf of St. Lawrence and Newfoundland
- Habitat: Demersal in depths of 30–540 m; rock, sand and mud substrates; larvae are pelagic, but may be most common in the lower part of water column
- Spawning: Spring-summer in St. Lawrence River Estuary; Jul-Sep in Arctic

Eggs: – Undescribed

- Larvae: Hatching size undescribed
 - Preanal ventral outline tends to bulge (more so than in *L. coheni*)
 - Suction disk appears at 7.4 mm
 - Larger eye diameter to disk length ratio than in comparably sized L. coheni
 - Nostrils split into 2 pairs at 12.7-15.4 mm
 - Preanus length 35% SL during preflexion, increases to 43% SL during flexion, then decreases to 39% SL in postflexion
 - Snout to anal fin 35% SL during preflexion, increases to 45% SL during later stages
 - Head length 18% SL in preflexion, increases to 28% SL in postflexion (smaller than in L. coheni)
 - Flexion occurs between 7.4 and 10.3 mm
 - Sequence of fin ray formation: $P_1 D, A C$
 - Notch does not form in anterior part of dorsal fin; fin rays are uniform or graduated in length
 - Pigmentation includes a cluster of small melanophores on the medial surface of the pectoral fin base; in preflexion, larvae develop a row of melanophores along base of anal fin, on dorsolateral surface of gut, lateral melanophores on tail, and dorsal surface of head; all pigment more dense in Arctic larvae
- Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids
 - 2. Similarities to larvae of *L. coheni* and *L. tunicatus*:
 - High numbers of myomeres and fin rays
 - Large sizes at all developmental stages
 - No development of notch in anterior part of dorsal fin

Late Flexion Larva:



H. 20.7 mmNL (Arctic)

Figures:Adult: D. R. Harriott (Scott and Scott, 1988); A–H: M. P. Fahay (Able *et al.*, 1986)References:Able, 1978; Able and M^cAllister, 1980; Able *et al.*, 1986; Scott and Scott, 1988

Liparis gibbus





Liparis atlanticus (Jordan and Evermann, 1898) Liparidae

Atlantic sea snail



Meristic Characters			
Myomeres:	38–42		
Vertebrae:	38–42		
Dorsal fin rays:	31-35		
Anal fin rays:	25-29		
Pectoral fin rays:	27-31		
Pelvic fin rays:	Disk		
Caudal fin rays:	5-6+5-6		

- Range: Western North Atlantic Ocean from Ungava Bay to Connecticut and Long Island, including Georges Bank
- **Habitat**: Intertidal pools to subtidal, usually <2 m; rarely as deep as 90 m
- Spawning: Mar–Jun in New Hampshire, mostly Jun in Newfoundland and northern Gulf of St. Lawrence; larvae present Jun–Sep in St. Lawrence River Estuary

Eggs: – Undescribed

Larvae: - Hatching size undescribed; early larvae have large, rounded head, slender tapering tail

- Preanal ventral outline tends to be flat
- Suction disk becomes visible at 3.3 mmNL
- Smaller eye diameter to disk length ratio at comparable sizes than Liparis inquilinus
- Nostril splits into 2 pairs at 5.1-9.4 mmNL
- Preanus length increases during preflexion and flexion (39–50% SL), then decreases during postflexion (45% SL) as anus migrates anteriorly
- Snout to anal fin increases from 41% SL during preflexion to 53% SL during postflexion
- Head length increases from about 20% SL during preflexion to 28% SL during postflexion
- Flexion occurs between 5.8 and 17.1 mm
- Sequence of fin ray formation: P_1 , D, A C
- Notch forms in anterior part of dorsal fin as anterior fin rays elongate
- Anterior dorsal fin rays form fleshy, finger-like extensions during postflexion stage; become more pronounced in adults, especially males in spawning season
- During preflexion, larvae have a row of melanophores along base of anal fin and on dorsolateral surface of gut; lateral melanophores form on tail during preflexion stage (soon after hatching); spots form a band across midpoint of body, expand anteriorly and posteriorly; a cluster of small melanophores on the medial surface of the pectoral fin base; scattered spots on head, especially ventral surface; melanophores on dorsum of head form later than in *L. gibbus* or *L. coheni*; small melanophores on venter between disk and anus highly variable, may be absent
- Entire head and body become covered with melanophores during postflexion stage
- Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids
 - 2. Similarities to larvae of *L. inquilinus*:
 - Low numbers of myomeres and fin rays
 - Small sizes at all developmental stages
 - Development of notch in anterior part of dorsal fin

Figures: Adult: D. R. Harriott (Scott and Scott, 1988); A–B, E: M. P. Fahay (Able *et al.*, 1986); C–D: Betsy Washington (Able *et al.*, 1986)

References: Detwyler, 1963; Able, 1978; Able and M^cAllister, 1980; Able and Irion, 1985; Able et al., 1986

Liparis atlanticus



E. 9.2 mmNL

Liparis inquilinus Able, 1973 Liparidae Inquiline snailfish



Range:	Western North Atlantic Ocean from Newfoundland to Cape Hatteras, includ-				
Habitat:	Demersal in depths of 3–97 m; often commensal with sea scallop (<i>Placopecten magellanicus</i>), especially during summer; leave scallop association and move into shallower water to spawn during fall and winter	Meristic Characters Myomeres: 38–42 Vertebrae: 38–42 Dorsal fin rays: 33–38			
Spawning:	Feb–Apr; larvae planktonic Apr–Jun; eggs deposited in clumps, possibly on hydroids; individual females spawn more than one batch	Anal fin rays: 28–31 Pectoral fin rays: 30–35 Pelvic fin rays: Disk Caudal fin rays: 5+5–6			
Eggs:	 Demersal, adhesive, pinkish Diameter: 1.0–1.3 mm Oil globules present: single or 3–8 				
Larvae:	 Hatching size undescribed; early larvae have large, rounded head, slender tapering tail Preanal ventral outline tends to bulge, with disk directed sightly anteriad Larger eye diameter to disk length ratio at comparable sizes than <i>Liparis atlanticus</i> Suction disk becomes visible at 3.7–6.0 mm Preanus length increases during preflexion and flexion (42–45% SL), then decreases during postflexion (40% SL) as anus migrates anteriorly Snout to anal fin increases from 41% SL during preflexion to 46% SL during postflexion Head length increases from about 21% SL during preflexion to 25% SL during postflexion Flexion occurs between 8.1–19.0 mm Sequence of fin ray formation: P₁, C, D, A Notch forms in anterior part of dorsal fin as anterior fin rays elongate during postflexion stage Anterior dorsal fin rays are elongate and resemble those of adult by 10.2 mmTL, but lack the fleshy extensions found in <i>L. atlanticus</i> Lateral melanophores on tail do not begin forming until flexion is well underway; melanophores begin in region of nape and expand posteriorly at about 8.0 mm; a cluster of small melanophores on the medial surface of the pectoral fin base; at preflexion, larvae have a row of melanophores along base of anal fin and on dorsolateral surface of gut; small melanophores appear behind chin and between disk and anus during flexion stage; dorsum of head unpigmented until about 12.0 mm; melanophores on pectoral fin rays variable, may be absent 				
Note:	1. See <i>Liparis tunicatus</i> page for summary of distinguishing characters in larva	e of liparids and cyclopterids			
	2. Similarities to larvae of <i>L. atlanticus</i> :				
	 Low numbers of myomeres and fin rays 				
	 Small sizes at all developmental stages 				
	 Development of notch in anterior part of dorsal fin 				

Liparis inquilinus



A. 4.9 mmNL



B. 8.5 mmNL



C. 8.5 mmNL (Ventral View)



D. 16.4 mmNL

Careproctus reinhardti (Krøyer, 1862) Liparidae



Range:	Arctic and North Atlantic oceans from Laptev Sea to Greenland and Gulf of St. Lawrence]
Habitat:	Demersal in depths of 200–1,000 m; larvae are also demersal (unlike pelagic larvae of other liparids)	Meristic Characte Myomeres: Vertebrae:	ers 56–63 56–63
Spawning:	Undescribed; small individuals (<20 mm) have been collected throughout spring and summer in St. Lawrence River Estuary and Chaleur Bay	Dorsal fin rays: Anal fin rays: Pectoral fin rays:	50–58 41–52 25–34
Eggs:	– Undescribed	Pelvic fin rays:	Disk
Larvae:	 Hatch at advanced stage of development; size undescribed Suction disk present at hatching Adult complements of fin rays present at hatching Diagnostic meristic characters develop before hatching Nostrils do not split into 2 pairs as in other liparids Head length increases from 19% SL during flexion to 21.5% SL during postflexion Preanus length changes little from 27.5% SL during flexion to 25% SL during postflexion Snout to anal fin changes little; about 32% SL through development 16–30 pyloric caeca present (8–13 in a congener, <i>C. ranula</i>) Flexion complete by 17.2–21.1 mm Postflexion stage larvae resemble adults, except have narrower tails and lack elongate rays on lower lobe of pectoral fin Peritoneum only lightly pigmented; other pigment uniformly distributed over-all except that head and ventri surface of gut are unpigmented 		ver lobe of and ventral

Note: 1. See Liparis tunicatus page for summary of distinguishing characters in larvae of liparids and cyclopterids

Sea tadpole

Careproctus reinhardti



B. 12.6 mmSL (Ventral View) 1004

Paraliparis calidus Cohen, 1968 Liparidae Lowfin snailfish



Range:	Western North Atlantic Ocean from Gulf of St. Lawrence to Gulf of Mexico
Habitat:	Demersal in depths of 150-732 m; larvae probably occur very near the bot-
	tom

Spawning: Undescribed; small juveniles have been collected in ice-free months from May–Oct

Eggs: – Diameter: 2.9–3.2 mm

Larvae: - Hatching size undescribed, but probably hatch at advanced stage of development

- Suction disk lacking
- Dorsal and anal fin ray counts high; adult complements present at 14.0 mm
- Flexion complete by 18.6-20.7 mm
- Head length 13–17% SL in flexion and postflexion stages
- Preanus length 23 to 21% SL in flexion and postflexion stages
- Snout to an us about 30% SL
- Teeth in several rows (single row in P. copei)
- Nostrils do not split into 2 pairs as in other liparids; (inadvertently illustrated as 2 in Able et al., 1984)
- Pectoral fins bi-lobed; any rays in interspace less-developed than in P. copei
- Note 6 principal caudal fin rays (8 or 9 in P. copei)
- Pigment includes melanophores on dorsal surface of nape and along dorsal fin base; abdomen and gill chamber darkly pigmented; lacks dark pigment around mouth

Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids

- 2. Similarities to larvae of *P. copei*:
 - Lacks suction disk at all sizes
 - High dorsal and anal fin rays counts
 - Pectoral fin rays in 2 groups with rudimentary rays in interspace
 - Darkly pigmented peritoneum
 - Anterior position of anus

Meristic Characters			
Myomeres:	_		
Vertebrae:	_		
Dorsal fin rays:	58-63		
Anal fin rays:	54-58		
Pectoral fin rays:	20-22		
Pelvic fin rays:	none		
Caudal fin rays:	6		

Paraliparis calidus



A. 12.9 mmSL



B. 12.9 mmSL (Ventral View) **Paraliparis copei** Goode and Bean, 1896 Liparidae Blacksnout snailfish



Myomeres: Vertebrae:

Dorsal fin rays:

Pectoral fin rays:

Anal fin rays:

Pelvic fin rays:

Caudal fin rays:

Meristic Characters

59-68

54 - 60

20-22

none

8-9

 Range:
 North Atlantic Ocean and eastern South Atlantic Ocean; in the western North Atlantic from Davis Strait to Cape Hatteras

Habitat: Demersal in depths of 200–1,088 m

Spawning: Undescribed; possibly year-round off Virginia; possibly spring-summer in St. Lawrence River Estuary

Eggs: – Undescribed

Larvae: - Hatching size undescribed, but probably hatch at advanced stage of development

- Suction disk lacking
- Dorsal and anal fin ray counts high; adult complements present at small size
- Flexion size undescribed
- Head length about 16.5% SL
- Preanus length about 23% SL
- Snout to anal fin about 30% SL
- Teeth in single row (several rows in *P. calidus*)
- Nostrils do not split into 2 pairs as in other liparids; (inadvertently illustrated as 2 in Able et al., 1984)
- Pectoral fins bi-lobed; any rays in interspace better-developed than in P. calidus
- Teeth in single row (several rows in *P. calidus*)
- Note 8 or 9 principal caudal fin rays (6 in P. calidus)
- Peritoneum, gill chamber darkly pigmented; dark pigment present around mouth and on snout; pigment absent on nape and along dorsal fin base
- Note: 1. See *Liparis tunicatus* page for summary of distinguishing characters in larvae of liparids and cyclopterids
 - 2. Similarities to larvae of *P. calidus*:
 - Lacks suction disk at all sizes
 - High dorsal and anal fin ray counts
 - Pectoral fin rays in 2 groups with rudimentary ray in interspace
 - Darkly pigmented peritoneum
 - Anterior position of anus

Figures: Adult: Goode and Bean, 1896; A: Betsy Washington (Able *et al.*, 1986) References: Wenner, 1979; Able and Irion, 1985; Able *et al.*, 1986

Paraliparis copei



A. 24.0 mmSL